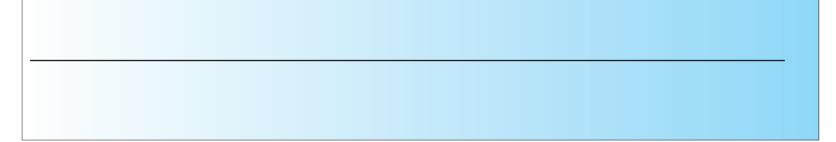
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AN EFFICIENT IN VITRO REGENERATION OF SHOOT ISBJ FROM COTYLEDON DERIVED CALLUS OF PIGEON PEA (CAJANUS CAJAN (L.) MILLSP.)

fB

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Abstract:-Pigeon pea (*Cajanus cajan* (L.) Millsp.) is an important grain legume of the semi-arid tropics. It provides protein rich food. Pigeon pea being recalcitrant species, regeneration of multiple shoots via callus induction and organogenesis was achieved from cotyledon explants of pigeon pea. Callus induction and shoot regeneration at various frequencies were observed using different concentration and combination of growth regulators. The highest callus formation was observed on MS medium + 1.0 mg l IAA + 0.9 mg l Kinetin. The Highest shoot formation was obtained on MS medium fortified with 2.0 mg l⁻¹ BAP + 0.2 mg l¹ NAA and 0.4 mg l¹ GA, Regenerated plants were successfully established in the soil after acclimatization.

Keywords: Cajanus cajan (L.) Millsp.), in vitro regeneration, Cotyledon explants.

1. INTRODUCTION

Pigeon pea or red gram (*Cajanus cajan* L. Millsp.) is one of the major grain legumes of the tropics and subtropics. Pigeon pea is a high protein grain legume and caters the protein requirement of the majority of the population in the Indian subcontinent (George and Eapen, 1994). It is a source of protein for 80% vegetarian in India. In India, red gram is prone to more than 200 species of insects (Anonymous, 1978), among which pod borer (*Helicoverpa armigera*) causes enormous losses (46.6 - 63.6%). *Helicoverpa armigera* is a polyphagous lepidopteron insect, present throughout the year and can complete up to seven generations in a year under favorable conditions. The demand for Pigeon pea boosts the production of high yielding and disease resistant varieties in India and the world market. To meet the increasing demand for these crops *In vitro* tissue culture followed by gene transfer would be an efficient and economical method to obtain a large number of disease free plants within short span of time.

Legumes are one of the most important groups of crop plants and have been subjected of efforts to improve desirable traits including their *in vitro* culture response. Since legumes are notoriously recalcitrant to regenerate from tissue culture, much effort has been devoted to developing and optimizing efficiencies in vitro regeneration system to facilitate a variety of technologies (Murashige and Skoog 1962). In Pigeon pea attempts to regenerate plants from various explants have been attempted, these include leaves (Dayal and Lavanya, 2003; Eapen and George, 1993; Eapen *et.al.*,1998; Geetha *et.al.*,1999; Singh *et.al.*,2002) cotyledonary node (Frankalin *et.al.*,1998; Geetha *et.al.*,1999; Mohan & Krishnamurthy, 1999; Shiva Prakash *et.al.*,1994; Singh *et.al.*,2002), epicotyls (George and Eapen, 1994) and shoot apics (Singh *et.al.*,2002; Singh *et.al.*,2004).

The Pigeon pea is recalcitrant in its regeneration potential, which becomes difficult to apply genetic transformation techniques. Therefore in the present research study, an attempt has been made to develop an efficient regeneration protocol by using cotyledon explants of pigeon pea.

2. MATERIALS AND METHODS

Collection of Plant Material

Certified Seeds of (*Cajanus cajan* L. Millisp.) Co-6 cultivar were obtained from the Centre for Plant breeding & Genetics, Tamil Nadu Agriculture University, Coimbatore.

M. Prabhakaran and S. Elumalai , "AN EFFICIENT *IN VITRO* REGENERATION OF SHOOT FROM COTYLEDON DERIVED CALLUS OF PIGEON PEA (*CAJANUS CAJAN* (L.) MILLSP.) " Indian Streams Research Journal | Volume 3 | Issue 12 | Jan 2014 | Online & Print

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Explant Preparation

Mature surface sterilized seeds cultured directly on MS basal agar medium without any growth regulators. Cotyledon explants obtained from the plant after 7 days and are surface sterilized using the following procedure (Santombi & Sharma, 2007)

Soaked the explants in bavistin solution f or exactly 15 minutes Washed the explants with tap water 3 - 4 times Washed the explants with double distilled water twice Washed the explants with Tween 20 and Sodium hypochlorite for 4 minutes Washed the explants with autoclaved double distilled water 4 times Washed the explants with autoclaved double distilled water 4 times Washed the explants with 70 % alcohol for exactly 30 seconds Immediately washed the explants with Hgcl₂ for exactly 2 and a half minutes Washed the explants with double distilled water 4 times to remove traces of ethanol and HgCl₂

Culture Media and Conditions

Cotyledon explants were transferred to regeneration media containing MS salts B_s vitamins⁵ and 3% sucrose and the pH of the medium is adjusted to 5.6 (standard pH for MS medium). MS medium with 1.0 mg/AA + 0.9 mg/Kinetin was used in the induction of callus and MS medium with 2.0 mg 1⁻¹ BAP + 0.2 mg 1⁻¹ NAA and 0.4 mg 1⁻¹ GA₃was used in the shoot induction. The explants were transferred and sub cultured in fresh medium in every three weeks and regenerated callus were transferred to shooting media.

Light

The light source is provided by fluorescent lamps and the photoperiod is controlled by an hourly timer. The fluorescent lamps produce better quality of light and which is distributed uniformly and generates less heat. The pigeon pea requires an illumination of around 2000 - 3000 Lux. The inoculated explants are maintained less than 16 hours photoperiod light for enhanced growth.

Acclimatization

Healthy plants with 5 - 7 cm long with different ages (15, 22 and 29 days) of rooted shoots were individually removed from the culture jars, and their roots washed carefully with tap water and were transferred to pots containing soil, soil with sand (1:1) and soil with compost (1:1) for observation on survivability of platelets under ex vitro condition.

Statistical Analysis

The results obtained from the growth observation of the plants grown in different medium with cotyledon explants were statistically analyzed using a stat pack version - 4.01 (161209) copy written by Berty Edwin. The results are interpreted based on the T - Test representing the P - value along with the standard error of each sample. Through this analysis, P - value interprets the significant growth of the explants grown on a different medium.

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3. RESULTS AND DISCUSSION

Surface sterilized seeds, cultured directly on MS basal agar medium without any growth regulators in the phyta jar which showed 40 - 50% germination in 5 days; 95 - 100 % germination was observed after 7 days (Fig 1). This seedling plant cotyledon explants (excised from 7 days old seedlings), were used for morphogenic response. Callus induction was observed onto MS and B media containing different concentration and combination of IAA, Kinetin, 2, 4-D, NAA and BAP within 8 -15 days of incubation of cotyledon explants depending upon the concentration and combination of hormones. Callus induction noticed in all media formulations. But there was a wide range of variation in average fresh weight and color. The highest callus induction was observed on MS medium containing 1.0 mg 1 IAA + 0.9 mg l¹ Kinetin (Fig. 2; Table 1, 1a). The highest callus growth in terms of fresh weight (0.802 g) was observed and color of calli was mostly whitish green and light green. It was observed only light green calli produce shoot buds. Proliferation of shoot buds was observed in MS + 2.0 mg 1 NAA and 0.4 mg 1 GA₃(Fig. 2, Table 2, 2a). The shoot buds first appeared as nodular growth within 3-4 weeks of culture and at the end of 4 weeks this nodular growth increased in size and produced leaf primordial. The highest shoot induction in terms of height of shoots. Maximum number of shoot buds were obtained in MS + 2.0 mg l¹ NAA and 0.4 mg l¹ GA₃ Root formation as recorded on MS + 1.0 mg l⁻¹ IBA and 0.5 mg l⁻¹ NAA (Fig. 2). Regenerated plants were successfully established in the soil.

The Pigeon pea *in vitro* regeneration depends on several factors like concentration of growth hormones, composition of media, media pH, types of explants, photoperiod time and light condition. In this study, cotyledons used as a explants, while this confirms previous reports



5 days old seedling plants





3

7 days old seedling plants

'An Efficient In Vitro Regeneration Of Shoot From Cotyledon Derived Callus Of Pigeon Pea (cajanus Cajan (l.) Millsp.)

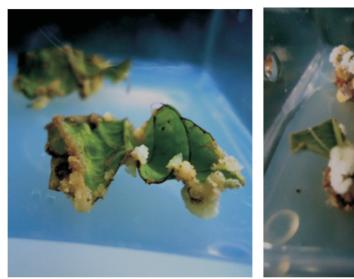


Fig A. 15 days' old regeneration



Fig B. 25 days' old regeneration



Fig C. 35 days' old regeneration



Fig D. 45 days' old regenerati

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Fig 2. Cotyledon as explants on (A8) regeneration media, callus $-1.0 \text{ mg } \Gamma^1 \text{ IAA} + 0.9 \text{ mg } \Gamma^1 \text{ Kinetin, shoot} - 2.0 \text{ mg } \Gamma^1 \text{ BAP} + 0.2 \text{ mg } \Gamma^1 \text{ NAA} + 0.4 \text{ mg } \Gamma^1 \text{ GA, root} - 1.0 \text{ mg } \Gamma^1 \text{ IBA} + 0.5 \text{ mg } \Gamma^1 \text{ NAA}$

 Table 1— Regeneration of callus induction medium for Pigeon pea (*Cajanus cajan* L. Millsp.) *Co - 6*. Three combinations of medium responded well and A8 media responded maximum growth

 Growth regulators
 Explant type

 (mg/l)
 8 days 12 days 15 days

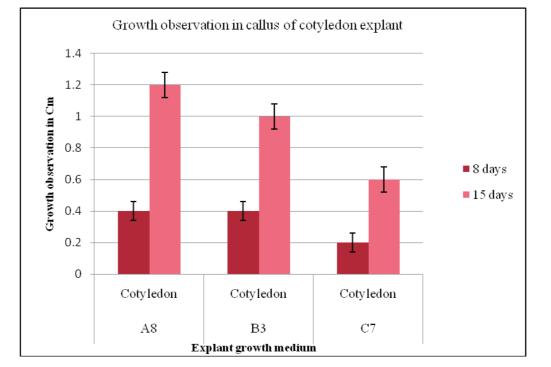
A8 – 1.0 mg ⁻¹ IAA +		A8	0.4	0.8	1.2
0.9 mg l ⁱ KIN	cotyledon	B3	0.4	0.6	1.0
B3 –1.0 mg l ¹ 2,4-D		C7	0.2	0.4	0.6
C7 – 1.0 mg 1 KIN					
AQ D2 C7 Different		ad different and	. 1. in ation of	£ 1 1 C	1:-

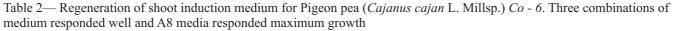
A8, B3, C7 – Different concentration and different combination of MS media IAA – Indole Acetic Acid, KIN – Kinetin, 2, 4-D – 2,4 - Dichlorophenoxy acetic acid

'An Efficient In Vitro Regeneration Of Shoot From Cotyledon Derived Callus Of Pigeon Pea (cajanus Cajan (l.) Millsp.)

Table 1a— Statistical growth observation analysis of cotyledon explants

OUTPUT / RESULT PANEL							
Treatments	n	Mean	SE	T-test	P-value	Result	
8 days	3	0.3333	0.0667	5.1962	0.0351	*	** - P < 0.01
15 days	3	0.9333	0.1764				* - P < 0.05
							NS - Not Significant





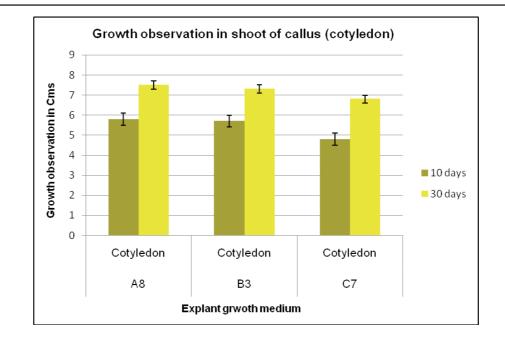
Growth regulators (mg/l)	Explant type	Medium number			on (Cm) s 30 days
$(\Pi \mathcal{G}^{(1)})$			10 duys	20 auy	, 50 auys
A8 – 2.0 mg ⁻¹ BAP +		A8	5.8	6.9	7.5
0.2 mg l ⁱ NAA	cotyledon	B3	5.7	6.7	7.3
0.4 mg l ⁱ GA3		C7	4.8	5.9	6.8
$B3 - 2.0 \text{ mg } 1^{l} BAP +$					
0.5 mg l ⁱ NAA					
C7 – 2.0 mg 1 KIN+					
1.0 mg l ¹ BAP					
A8, B3, C7 – Different	concentration	and different comb	ination of	MS med	lia
DAD D. 1.	TAA D	T 1 1		0:11	

BAP - Benzyl amino purine, NAA - Napathalene acetic acid, GA3- Gibberellic acid, KIN - Kinetin,

Table 2a— Statistical growth observation analysis in the shoot of cotyledon explant

OUTPUT / RESULT PANEL							
Treatments	n	Mean	SE	T-test	P-value	Result	
10 days	3	5.4333	0.3180	14.6995	0.0046	**	** - P < 0.01
30 days	3	7.2000	0.2082				* - P < 0.05
							NS - Not Significan

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An Efficient In Vitro Regeneration Of Shoot From Cotyledon Derived Callus Of Pigeon Pea (cajanus Cajan (l.) Millsp.)

Cotyledons (Frankalin *et.al.*,1998; Geetha *et.al.*, 1999; Mohan and Krishnamurthy, 1998; Shiva Prakash *et.al.*,1994; Singh *et.al.*, 2004), which can be induced to differentiate into adventious callus and shoots that can be used for efficient production of transgenic Pigeon pea.

The combination of BAP and NAA increased the number of multiple shoots, while this confirms previous results, regeneration of pigeon pea resulted best with MS medium supplemented with 2.0 mg¹ BAP + 0.2 mg¹ NAA + 0.4 mg¹ GA₃ (Dayal *et.al.*, 2003; Eapen and George 1993; Eapen *et.al.*, 1998; Frankalin *et.al.*, 1998; Geetha *et.al.*, 1999; George and Eapen 1994: Mohan and Krishnamurthy, 1998; Shiva Prakash *et.al.*, 1994; Singh *et.al.*, 2004).

In the present investigation the dose of BAP and NAA is known to be produced critical multiple shoot organogenesis. Therefore we compared the response of various concentrations of BAP and NAA. The maximum number of multiple shoot buds were obtained in MS + 2.0 mg⁻¹ BAP + 0.2 mg⁻¹ NAA and 0.4 mg⁻¹ GA₃

4. ACKNOWLEDGEMENT

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5. REFERENCES

1. Anonymous, A summary of high-yield lac production. Research group for high-yield lac production, Developing Status of Lac Production, 2 (1978) 1-4.

2.Dayal S, Lavanya M, Devi P & Sharma K K, An efficient protocol for shoot regeneration and genetic transformation of pigeon pea (Cajanus cajan [L.] Millsp.) using leaf explants, Plant Cell Rep, 21 (2003)1072-1079.

3.Eapen S & George L, Plant regeneration from leaf discs of peanut and pigeon pea: influence of benzyladenine, indoleacetic acid and indoleacetic acid-amino acid conjugates, Plant Cell Tissue Organ Cult, 35 (1993) 223-227.

4.Eapen S, Tivarekar S & George L, Thidiazuron-induced shoot regeneration in pigeon pea (Cajanus cajan L.), Plant Cell Tissue Organ Cult, 53 (1998) 217-220.

5.Ebida A I A & Hu C, In vitro morphogenic responses and plant regeneration from pepper (Capsicum annum L.cv Early California Wonder) seedling explants, Plant Cell Rep, 13 (1993) 107-110.

6.Frankalin G, Jayachandran R, Melehias G & Ignacimuthu S, Multiple shoot induction and regeneration of pigeon pea (Cajanus cajan (L.) Millsp.) cv. Vamban 1 from apical and axillary meristem, Curr. Sci, 74 (1998) 936-937.

7.Geetha N, Venkatachalam P & LakshmiSita G, Agrobacterium - mediated genetic transformation of pigeon pea (Cajanus cajan L.) and development of transgenic plants via direct organogenesis, Plant Biotechnol, 16 (1999) 213-218.

8.George L & Eapen S, Organogenesis and embryogenesis from diverse explants in pigeon pea. Plant Cell Rep, 13 (1994) 417-

420. 9.Mohan M L & Krishnamurthy K V, Plant regeneration in pigeon pea (Cajanus cajan (L.) Millsp.) by organogenesis, Plant Cell Rep 17 (1998) 705-710.

10.Murashige T & Skoog F, A revised medium for rapid growth and bioassays with tobacco tissue cultures, Physiol Plant, 15

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'An Efficient In Vitro Regeneration Of Shoot From Cotyledon Derived Callus Of Pigeon Pea (cajanus Cajan (l.) Millsp.)

11.Polanco M C & Ruiz M L, Plant Cell Rep; 17 (1997) 22-26. 12.Santombi K & Sharma G J, Micropropogation of Capsicum frutscens L. using axillary shoot explants, Sci Hortic Amsterdam, 13 (2007) 96-99.

13.Shiva Prakash N, Pental D & Bhalla-Sarin N, Regeneration of pigeon pea (Cajanus cajan) from cotyledonary node via multiple shoot formation, Plant Cell Rep, 13 (1994) 623-627.
14.Singh N D, Sahoo L, Saini R, Sarin N B & Jaiwal P K, In vitro shoot organogenesis and plant regeneration from

cotyledonary node and leaf explants of pigeon pea (Cajanus cajan L. Millsp.), Physiol. Mol. Biol. Plant, 8 (2002) 133-140. 15.Singh P K, Mithilesh Kumar C P, Chaturvedi D, Yadav R & Tuli, Development of a hybrid d-endotoxin and its expression in tobacco and cotton for control of a polyphagous pest Spodoptera litura, Transgenic Res, 13 (2004) 397-410.

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